

# Certificate Of Analysis



## Client:

### Particle Peptides

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## Sample Identification

Sample Name	MOTS-C 10 mg	Batch Number	2025259	Date Published	2025-07-25 18:22
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## Results for Lyo-0055

Analysis of Peptide Identity, Content and Purity	Result	Unit	Uncertainty	Reporting Limit
MOTS-C Assay Peptide Screening	9.94	mg	[± 0.05]	
MOTS-C Identification by RT Peptide Screening	0.999		[± 0.005]	
MOTS-C Identification by spectrum Peptide Screening	1.00		[± 0.01]	
MOTS-C Purity Peptide Screening	> 99.8	%		


Bioburden	Result	Unit	Uncertainty	Reporting Limit
Total Aerobic Microbial Count USP <61> Plate Count Method	25	CFU/g	[± 2]	>= 1000
Total Yeast and Mold Count USP <61> Plate Count Method	Not detected	CFU/g		>= 100

Endotoxin Analysis	Result	Unit	Uncertainty	Reporting Limit
Bacterial Endotoxin USP<85> Bacterial Endotoxin Chromogenic Test	0.0100	EU/mg	[± 0.0002]	> 0.5

Heavy Metals	Result	Unit	Uncertainty	Reporting Limit
Arsenic Elemental Impurities Screening	Not detected	ppm		>= 1.5
Cadmium Elemental Impurities Screening	Not detected	ppm		>= 0.5
Cobalt Elemental Impurities Screening	Not detected	ppm		>= 25
Lead Elemental Impurities Screening	Not detected	ppm		>= 1.5

Heavy Metals	Result	Unit	Uncertainty	Reporting Limit	
Nickel Elemental Impurities Screening	Not detected	ppm		>= 25	△
Quicksilver Elemental Impurities Screening	Not detected	ppm		>= 1.5	△
Vanadium Elemental Impurities Screening	Not detected	ppm		>= 25	△

# Attachments for Lyo-0055

	<b>Method Specification</b>		
Determination of identity, content and purity of MOTS-C			
Document number <i>MOTS_001_2025</i>	Superseded document -	Number of pages 2	

## 1. Content and Purity Assessment

### 1.1. Instrumentation

Module	Name	Serial Number
System Controller	Shimadzu SCL-10ADvp	C21014112659
Degassing Unit	Shimadzu DGU-14A	NA
LPGE valve	Shimadzu FCV-10Avp	NA
Pump	Shimadzu LC-10ADvp	C20964130094
Autosampler	Shimadzu SIL-10ADvp	C21054109114
Column Thermostat	Shimadzu CTO-10ACvp	C21033770144
Detector	Shimadzu SPD-10ADvp	C20994233588

### 1.2. Chromatographic conditions

Chromatographic conditions	
Eluent A	1% TFA in Water (HPLC, Gradient Grade)
Eluent B	1% TFA in Acetonitrile (HPLC, Gradient Grade)
Flow rate	1mL/min
Program	Gradient elution
Injection volume	2 µL
Column Temperature	35°C
Column	Avantor ACE UltraCore C18, 150x3,0 mm, 3,5µm
Detection wavelength	214nm

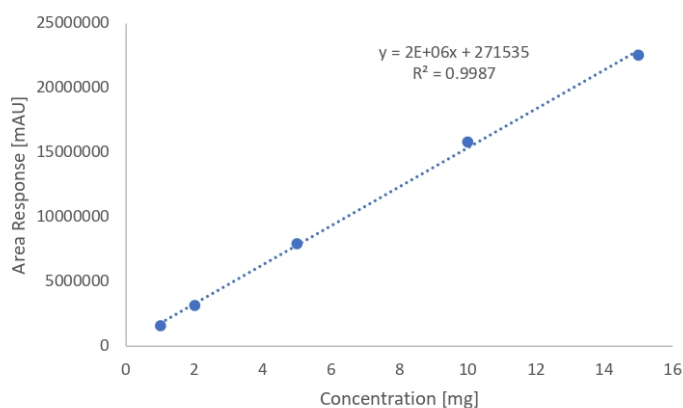
Gradient Program		
Time [min]	A [%]	B [%]
2	99	1
25	5	95
28	5	95
28.01	99	1
32	end	

### 1.3. Sample preparation

Whole amount of container was dissolved in 2mL of water (HPLC, Gradient Grade). Aliquote part of 1 mL was dispensed into HPLC vial for analysis.

### 1.4. Calibration curve

Calibration curve detail	
Quantitative method	External Standard
Calibration Type	Linear
Number of calibration points	5
Force through Zero	Disabled
Weighting Method	None



## 2. Purity assessment

### 1.5. Calculation of purity of peptide

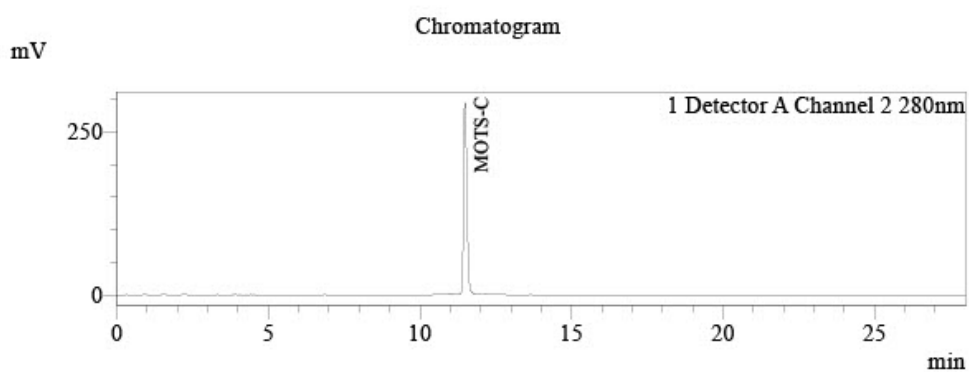
Purity of MOTS-C in sample was determined by area normalization method as ratio of MOTS-C peak and sum of all peaks detected at wavelength 214 nm.

$$Purity(\%) = \frac{Peak\ Area_{DSIP}}{\sum Peak\ Area_i}$$

# Analysis Report



Sample Information  
Injection Volume : 2  
Data File : LYO-0055-P01\_002\_004.lcd  
Method File : Peptide screening\_new.lcm  
Date Acquired : 30.06.2025 17:05:03




Peak Table

Detector A Channel 2 280nm

Peak#	Name	Ret. Time	Conc.	Unit	Area%
1	MOTS-C	11,483	9,941	mg	100,000
Total			9,941		100,000

Attachment for Lyo-0055  
Filename: MOTS-C.jpg

	<b>Method Specification</b>	
<b>Determination of bacterial endotoxin content of lyophilized samples</b>		
<i>Document number</i> <b>ENDOTOX_001_2025</b>	<i>Superseded document</i> -	<i>Number of pages</i> 2

## 1. Chromgenic LAL Assay Determination of Bacterial Endotoxin content of sample

### 1.1. Instrumentation

- Pipette set 1-1000 µL
- Thermostatically controlled water bath
- UV VIS spectrometer ( Shimadzu UV-1601)
- GenScript ToxinSensor Chromgenic LAL Endotoxin Assay kit

### 1.2. Chemicals

- LAL Reagent water (endotoxin free)
- Limulus Amoebocyte Lysate
- LAL Substrate
- Color Stabilizer #1
- Color Stabilizer #2
- Color Stabilizer #3
- 35% HCl (p.a.)

### 1.3. Sample preparation

1. Sample container was weighed prior to dissolution and measured weight was marked.
2. Sample was completely dissolved in its container by 2 mL of LAL Reagent water.
3. 100 µL of the sample was aliquoted for analysis.
4. After analysis container was emptied and dried.
5. Dry mass of container was measured and exact weight of dissolved content was determined as:

$$m_{dc} = m_{sample} - m_{container}$$

### 1.4. Toxin sensor Chromgenic LAL Endotoxin Assay kit preparation

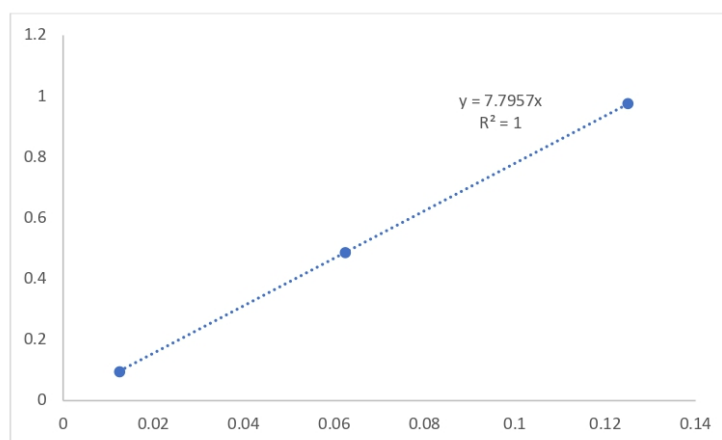
Procedures regarding preparation of reaction solutions possible to find in:

[https://www.genscript.com/site2/document/5292\\_20080806231827.PDF](https://www.genscript.com/site2/document/5292_20080806231827.PDF)

### 1.5. Measurement procedure

	Standards	Samples	Blank
Standards (mL)	0.1	-	-
Samples (mL)	-	0.1	-
LAL Reagent Water (mL)	-	-	0.1
LAL Solution (mL)	0.1	0.1	0.1
Mix well and incubate at 37°C for 27 min			
Substrate solution (mL)	0.1	0.1	0.1
Mix well and incubate at 37°C for 6 min			
Color Stabilizer #1 solution	0.5	0.5	0.5
Color Stabilizer #2 solution	0.5	0.5	0.5
Color Stabilizer #3 solution	0.5	0.5	0.5
Mix well and read the absorbance at 545nm			

### 1.6. Calibration curve



### 1.7. Calculation of endotoxin content

Endotoxin content of the sample was calculated from the calibration curve as:


$$Endotox[EU/mg] = \frac{\left(\frac{ABS_{sample}}{S_{calib}}\right) * 20}{m_{sample}}$$

$ABS_{sample}$  = Measured absorbance of sample

$S_{calib}$  = Slope of calibration curve

$m_{sample}$  = real measured mass of sample

20 = dilution factor of measured sample

	<b>Method Specification</b>		
Determination of bioburden of lyophilized samples			
Document number MIC_001_2025	Superseded document -	Number of pages 2	

## 1. Instrumentation and chemicals

### 1.1. Instruments used

- Sterile Syringe 2mL Luer
- Sterile needles
- Ready made PCA Plate ROTI Aquatest
- Ready made Sab4 Plate ROTI Aquatest

### 1.2. Chemicals

Sterile physiological solution (0.9% NaCl)

## 2. Sample preparation and inoculation

### 2.1 Sample preparation

1. Fresh sterile needle and syringe was used for measuring exactly 2 mL of sterile physiological solution.
2. Needle was changed and by new needle rubber top of peptide container was penetrated and 2 mL of sterile physiological solution was dispensed.
3. Content of container was completely dissolved and left for 5 minutes to settle potentially created bubbles.
4. This procedure is repeated for two vials.

### 2.2 Total Aerobic microbial count inoculation and cultivation

1. By sterile needle 1 mL of solution was filled into the sterile syringe.
2. Needle was placed above the flame for few seconds to sterilize.
3. Consequently 1 mL of solution was poured into the ready to use sterile petri dish filled with PCA agar and petri dish was closed.
4. Proces was repeated for two petri dishes.
5. With sterile needle, 1 mL of sterile physiological solution was filled into the sterile needle and was inoculated onto one sterile petri dish filled with PCA agar as negative control sample.
6. Samples and negative control sample were placed in incubator at temperature 37°C for 120h.

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Attachment for Lyo-0055  
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### 2.3 Total Yeast and Mold count inoculation and cultivation

1. By sterile needle 1 mL of solution was filled into the sterile syringe.
2. Needle was placed above the flame for few seconds to sterilize.
3. Consequently 1 mL of solution was poured into the ready to use sterile petri dish filled with Sab4 agar and petri dish was closed.
4. Proces was repeated for two petri dishes.
5. With sterile needle, 1 mL of sterile physiological solution was filled into the sterile needle and was inoculated onto one sterile petri dish filled with Sab4 agar as negative control sample.
6. Samples and negative control sample were placed in incubator at temperature 25°C for 72h.

## 3. Evaluation of results

After incubation time, colonies are counted as cfu (colonies forming units) and result per 1g of sample is determined as:

$$CFU_{avg} = \frac{\sum CFU_n}{n}$$

$CFU_{avg}$  = average CFU counted form  $n$  inoculations

$CFU_n$  = CFU counted per inoculation

$n$  = number of inoculations

$$CFU \text{ per gram} = \frac{CFU_{avg}}{m_s} * DF$$

$CFU_{avg}$  = Average CFU counted from  $n$  inoculations

$m_s$  = mass of sample (mg)

$DF$  = Dilution factor

If negative control sample is evaluated as positive, process have to be repeated due to possible contamination in the process of inoculation or incubation.

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Attachment for Lyo-0055  
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## Responsibles



**Mr. Ján Galbavý**  
Founder/Manager

Analysis results relate only to the samples tested.

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